



The ARRIVE guidelines 2.0: author checklist

The ARRIVE Essential 10

These items are the basic minimum to include in a manuscript. Without this information, readers and reviewers cannot assess the reliability of the findings.

Item	Recommendation	Section/line number, or reason for not reporting
Study design	1 For each experiment, provide brief details of study design including: a. The groups being compared, including control groups. If no control group has been used, the rationale should be stated. b. The experimental unit (e.g. a single animal, litter, or cage of animals).	Sec. 2.4: Sham; CLP; CLP+DEX; CLP+DEX+ML385. Suppl. Fig. 1 controls not fully described. Single mouse.
Sample size	2 a. Specify the exact number of experimental units allocated to each group, and the total number in each experiment. Also indicate the total number of animals used. b. Explain how the sample size was decided. Provide details of any <i>a priori</i> sample size calculation, if done.	Main study: 3/group x 4 = 12 mice (Sec. 2.27; Figs. 1-4). Suppl. Fig. 2: n=10; group basis/total unclear. No <i>a priori</i> calculation or sample-size rationale reported; n=3 limitation and post-hoc effect sizes noted.
Inclusion and exclusion criteria	3 a. Describe any criteria used for including and excluding animals (or experimental units) during the experiment, and data points during the analysis. Specify if these criteria were established <i>a priori</i> . If no criteria were set, state this explicitly. b. For each experimental group, report any animals, experimental units or data points not included in the analysis and explain why. If there were no exclusions, state so. c. For each analysis, report the exact value of <i>n</i> in each experimental group.	Not reported: no prespecified inclusion/exclusion criteria. Not reported whether any animals/data were excluded. Main analyses: n=3/group (Figs. 1-4). Suppl. Fig. 2: n=10; group basis unclear.
Randomisation	4 a. State whether randomisation was used to allocate experimental units to control and treatment groups. If done, provide the method used to generate the randomisation sequence. b. Describe the strategy used to minimise potential confounders such as the order of treatments and measurements, or animal/cage location. If confounders were not controlled, state this explicitly.	Random number table (Sec. 2.4). Confounder-control strategy (order/cage location) not reported.
Blinding	5 Describe who was aware of the group allocation at the different stages of the experiment (during the allocation, the conduct of the experiment, the outcome assessment, and the data analysis).	Blinded assessment: histology, IHC, TEM, and flow gating (Secs. 2.9-2.11, 2.23). Allocation, conduct, and overall analysis blinding not reported.
Outcome measures	6 a. Clearly define all outcome measures assessed (e.g. cell death, molecular markers, or behavioural changes). b. For hypothesis-testing studies, specify the primary outcome measure, i.e. the outcome measure that was used to determine the sample size.	Secs. 2.5, 2.9-2.23: W/D ratio, lung injury, inflammation, Nr2f2^H-1/GPX4, SOD/GSH/iron, TEM, macrophage phenotypes. No primary outcome for sample-size determination specified.
Statistical methods	7 a. Provide details of the statistical methods used for each analysis, including software used. b. Describe any methods used to assess whether the data met the assumptions of the statistical approach, and what was done if the assumptions were not met.	Sec. 2.27: Prism 10; mean +/- SD; one-way ANOVA/Tukey; Cohen's d; two-sided p<0.05. Normality assumed <i>a priori</i> ; n=3 prevented testing. No alternative method reported (Sec. 2.27; Limitations).
Experimental animals	8 a. Provide species-appropriate details of the animals used, including species, strain and substrain, sex, age or developmental stage, and, if relevant, weight. b. Provide further relevant information on the provenance of animals, health/immune status, genetic modification status, genotype, and any previous procedures.	Sec. 2.3: male C57BL/6; 6-8 weeks; 18-22 g. Sec. 2.3: CQMU Animal Center; SPF barrier; sterilized food/water ad libitum. Genotype/previous procedures not reported.
Experimental procedures	9 For each experimental group, including controls, describe the procedures in enough detail to allow others to replicate them, including: a. What was done, how it was done and what was used. b. When and how often. c. Where (including detail of any acclimatisation periods). d. Why (provide rationale for procedures).	Sec. 2.4: CLP/sham, isoflurane, PBS/DEX/ML385, 24-h endpoint, CO2 euthanasia, tissue collection. Sec. 2.4: ML385 -30 min; DEX/PBS +30 min; single dose; endpoint 24 h. Sec. 2.3: SPF barrier. Acclimatization not reported. Introduction/Sec. 2.4: CLP models sepsis-ALI; DEX protection; ML385 tests Nr2f2 dependence.
Results	10 For each experiment conducted, including independent replications, report: a. Summary/descriptive statistics for each experimental group, with a measure of variability where applicable (e.g. mean and SD, or median and range). b. If applicable, the effect size with a confidence interval.	Secs. 3.1-3.4; Figs. 1-4: mean +/- SD; n=3/group. Sec. 3.1: Cohen's d reported for selected endpoints; no confidence intervals.